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REFERENCES

ANSELL, G. B. & SPANNER, S. (1968). Biochem. J., 110, 201-206.
AQUILONIUS, S. M. & WINBLADH, B. (1972). Acta physiol. scand., 85, 78-90.
BARKER, L. A., DOWDALL, M. J. & WHITTAKER, V. P. (1972). Biochem. J., 130, 1063-1080.
CHAKRIN, L. W. & SHIDEMAN, F. E. (1968). Int. J. Neuropharmac., 7, 337-349.
DIAMOND, I. (1971). Arch. Neurol., 24, 333-339.
DOWDALL, M. J., BARKER, L. A. & WHITTAKER, V. P. (1972). Biochem. J., 130, 1081-1094.
SCHUBERTH, J., SPARF, B. & SUNDWALL, A. (1969). J. Neurochem., 16, 695-700.

Resistance of intestinal α -adrenoceptors to cold storage

It has been suggested that storage of rabbit intestine at $6-8^{\circ}$ for 24 to 72 h leads to selective impairment of α -adrenoceptor function (Lum, Kermani & Heilman, 1966; Lum, Heilman & Gaunt, 1967; Salimi, Kermani & others, 1970).

In the present experiments the α -agonist (—)-phenylephrine and the β -agonist (—)isoprenaline have been compared as inhibitors of the spontaneous isometric contractions in fresh and cold-stored segments of rabbit intestine, in the presence and absence of the selective antagonists, phentolamine and propranolol. The bathing solution used in the experiments was a modified Krebs solution with the following composition (mM):—NaCl 119.6, KCl 4.95, CaCl₂ 2.45, MgSO₄ 1.2, NaH₂PO₄ 1, sucrose 10, glucose 10.9, NaHCO₃ 25.

A complete dose-response curve for either agonist was determined on a fresh jejunal segment. One of the two antagonists was then added to the bathing solution and after an equilibration period of 30 min the dose-response curve determination for the agonist was repeated. A similar procedure was subsequently followed with an adjacent segment which was stored at $6-8^{\circ}$ for 24 h. The concentrations of antagonists which we used were those used in the experiments of Lum & others (1966). The potencies of the agonists have been expressed as pD₂ values (Ariens & van Rossum, 1957) where the maximal response was arbitrarily defined as complete extinction of at least one spontaneous contraction. The mean pD₂ values so obtained are shown in Table 1.

In fresh tissues, isoprenaline yielded log dose-response curves flatter than those produced by phenylephrine, the onset of its inhibitory action was slower and its effects lasted longer; however, its potency was similar to that of phenylephrine. These findings are in general agreement with those of van Rossum & Mujic (1965) and of Bowman & Hall (1970), and with the hypothesis of a dual adrenoceptor mechanism subserving inhibition in rabbit jejunum (Ahlquist, 1948; Ahlquist & Levy, 1959; Levy 1959; Furchgott, 1960).

Storage of tissues at $6-8^{\circ}$ for 24–48 h did not affect the rate of spontaneous movement (15 min⁻¹ for tissues equilibrated at 37°). In cold-stored tissues, the inhibitory potencies of both isoprenaline and phenylephrine were increased; there was less variability in tissue response; log dose-response curves were flatter; the onset and duration of the effects were indistinguishable from those observed in fresh tissues.

Phentolamine proved to be somewhat more potent an antagonist of phenylephrine in cold-stored tissues than in freshly excised tissues; thus the pA_2 (molar) estimates calculated from the results shown in Table 1 were 8.2 and 9.2 for fresh and cold-

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Agonist	Tissue condition	Mean pD₂ No antagonist	values \pm s.e. Phentolamine 100 ng ml ⁻¹	Propranolol 200 ng ml ⁻¹
(-)-Phenylephrine	Fresh	7.16 ± 0.29 (23)	5·35 ± 0·93 (7)*	7·97 ± 0·49 (15)
	Cold	7·97 ± 0·26 (15)	5·17 ± 0·18 (7)*	7·49 ± 0·24 (7)**
(-)-Isoprenaline	Fresh	7.66 ± 0.19 (35)	7·3 ± 0·47 (6)**	6.87 ± 0.19 (27)*
	Cold	8.07 ± 0.18 (25)	7·8 ± 0·46 (4)**	7·0 ± 0·17 (22)*

Table 1. Mean pD_2 values for phenylephrine and isoprenaline in the absence and presence of antagonists

() No of experiments.

* indicates that the value differs significantly (at P = 0.05) from appropriate control value on on the basis of an unpaired Student's t test.

** indicates that there was a significant difference (at P = 0.05) between these and the control values when a paired *t*-test was applied to values obtained on each preparation.

stored tissues respectively. Propranolol was a less potent antagonist of isoprenaline and was approximately equipotent in fresh and cold-stored tissues. The pA_2 (molar) estimates for propranolol/isoprenaline were 6.8 and 7.1 in fresh and cold-stored tissue respectively.

Some antagonism of isoprenaline by phentolamine and of phenylephrine by propranolol (the latter in cold-stored tissues only) was indicated when "within" tissue comparisons of pD_2 values were undertaken. Such findings suggest some overlap of the properties of α - and β -adrenoceptors in rabbit intestine, or more probably, that there may be some lack of specificity in the actions of the agonists and/or antagonists used. The results clearly indicate that cold storage of rabbit intestine under the present conditions does not lead to selective impairment of intestinal α -adrenoceptors. Possibly, the difference in the present results and those of Lum & others (1966, 1967) may relate to their use, and our omission of atropine from the bathing medium.

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REFERENCES

AHLQUIST, R. P. (1948). Am. J. Physiol., 153, 586-600.

AHLQUIST, R. P. & LEVY, B. (1959). J. Pharmac. exp. Ther., 127, 146-149.

ARIENS, E. J. & ROSSUM, J. M. VAN (1957). Archs int. Pharmacodyn. Thér., 110, 275-299.

BOWMAN, W. C. & HALL, M. T. (1970). Br. J. Pharmac., 38, 399-415.

 FURCHGOTT, R. F. (1960). In Ciba Foundation Symposium on Advenergic Mechanisms, Editors: Vane, J. R., Wolstenholm, G. E., and Conner, M. O., pp. 246-252. Boston: Little, Brown and Co. LEVY, B. (1959). J. Pharmac. exp. Ther., 127, 150-156.

LUM, B. K. B., KERMANI, M. H. & HEILMAN, R. D. (1966). Ibid., 154, 463-471.

LUM, B. K. B., HEILMAN, R. D. & GAUNT, M. A. (1967). Eur. J. Pharmac., 1, 109-113.

ROSSUM, J. M. VAN & MUJIC, M. (1965). Archs int. Pharmacodyn. Thér., 155, 418-431.

SALIMI, M., KERMANI, R. Z., DJAHANSOUZ, B. & GOLSHAN, Sh. (1970). Pharmacology, 4, 341-346.